Check List the journal of biodiversity data

ANNOTATED LIST OF SPECIES

Check List 14 (1): 213–224
https://doi.org/10.15560/14.1.213

PENSOFT

An updated list of sand flies (Diptera, Psychodidae, Phlebotominae) in the Federal District of Brazil

Aline Rapello,[1] Andrey José de Andrade,[2] Douglas de Almeida Rocha,[1] Jônatas C. B. Ferreira,[1] Renata Velôzo Timbó,[1] Marcos Takashi Obara,[1] Rodrigo Gurgel-Gonçalves[1]

1 Universidade de Brasília, Faculdade de Medicina, Área de Patologia, Laboratório de Parasitologia Médica e Biologia de Vetores, Campus Universitário Darcy Ribeiro, Asa Norte, 70910-900, Brasília, DF, Brazil. **2** Universidade Federal do Paraná, Setor de Ciências Biológicas, Departamento de Patologia Básica, Laboratório de Parasitologia Molecular, Jardim das Américas, 80050-540, Curitiba, PR, Brazil.
Corresponding author: Aline Rapello, aline_rapello@hotmail.com

Abstract
The Federal District of Brazil (FD) has the lowest Phlebotominae species richness in the country's Midwestern Region. Some of these species have been reported as *Leishmania* vectors. This study updates the list of sand flies in the FD by sampling 4 gallery forests. Using HP light traps and Shannon traps, 1,209 sand flies were captured and 16 species were identified. The most frequent species was *Bichromomyia flaviscutellata* (Mangabeira, 1942) ($N = 668$), followed by *Psathyromyia pradobarrientosi* (Le Pont, Matias, Martinez & Dujardin, 2004) ($N = 285$). *Brumptomyia guimaraesi* (Coutinho & Barretto, 1941), *Micropygomyia ferreirana* (Martins, Barretto & Pellegrino, 1956), *Pa. pradobarrientosi*, *Br. brumpti* (Larrousse, 1920), and *Evandromyia bourrouli* (Barretto & Coutinho, 1941) are reported for the first time in the FD. Sand flies richness in the FD (35 spp.) is relevant when compared to that of the state of Goiás (47 spp.), a state 59 times larger than the FD.

Key words
Psathyromyia pradobarrientosi; Brazilian savannah; species richness; list of species.

Academic editor: Kirstern Haseyama | Received 25 August 2016 | Accepted 25 August 2017 | Published 9 February 2018

Citation: Rapello A, Andrade AJ, Rocha DA, Ferreira JCB, Timbó RV, Obara MT, Gurgel-Gonçalves R (2018) An updated list of sand flies (Diptera, Psychodidae, Phlebotominae) in the Federal District of Brazil. Check List 14 (1): 213–224. https://doi.org/10.15560/14.1.213

Introduction

Phlebotomine sand flies (Diptera, Psychodidae, Phlebotominae) are insects with a wide geographic distribution where they are found in diverse climatic conditions and at a range of altitudes, as well as in rural, urban, and sylvatic environments. There are currently 1003 species, 537 of which are found in the Neotropical and Nearctic regions, and 277 in Brazil (Shimabukuro et al. 2017). Only 19 sand fly species have been incriminated as vectors of *Leishmania* species (Kinetoplastida, Trypanosomatidae), which may be etiological agents of cutaneous and visceral leishmaniasis (Shimabukuro and Galati 2011).

Most cases of leishmaniasis in the Americas originated in Brazil (Alvar et al. 2012). Between 2001 and 2012, approximately 3000 confirmed cases of visceral leishmaniasis (VL) and 50,000 of cutaneous leishmaniasis (CL) were reported in the Center-West Region of Brazil (http://www.saude.gov.br), where at least 127 phlebotomine species have been recorded (Almeida et al. 2015). This region is largely within the Brazilian Cerrado biome. In this savannah region, vast agricultural expansion, as well as occupation of the land due to unplanned urban growth may favor the occurrence of leishmaniasis (Galati et al. 1996, Carranza-Tamayo et al. 2010).

Copyright *Rapello et al.* This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

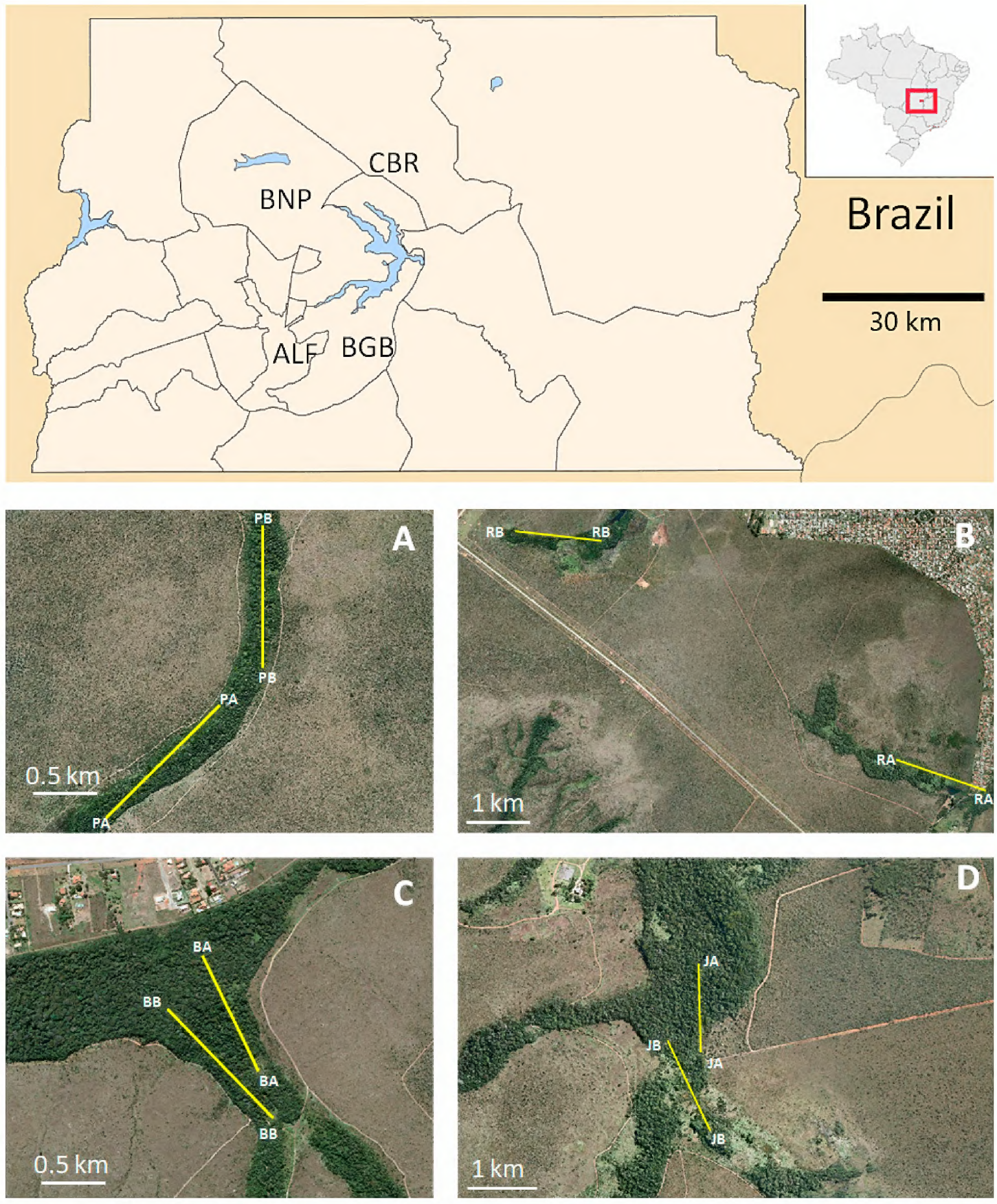

Figure 1. Geographic location of the four areas of the study performed in the Federal District of Brazil. **A.** BNP = Brasília National Park (trapping lines PA, PB). **B.** CBR = Contagem Biological Reserve (RA, RB). **C.** ALF = Água Limpa Farm within the University of Brasília (BA, BB). **D.** BGB = Brasília Botanical Garden (JA, JB). Source: CODEPLAN (Companhia de Planejamento do Distrito Federal2017 (http://ortofoto.mapa.codeplan.df.gov.br/demo/tms).

In the Distrito Federal do Brasil (Brazilian Federal District, hereinafter FD), there is evidence of autochthonous transmission of CL (Sampaio et al. 1980). The number of cases reported as originating from the FD in 2003, 2004, and 2005 were 31, 57, and 26, respectively (Sampaio et al. 2009). The first autochthonous case of VL in the FD occurred in 2005, and 3 deaths were confirmed by 2009 (Carranza-Tamayo et al. 2010). Between 2007 and 2012, 24 human cases of autochthonous VL were reported (data from Health Secretariat of the Federal District). In 2013, 98 suspect cases of VL were reported in the Brazilian National Notifiable Diseases Surveillance System (SINAN/NET), from which 47 were confirmed. Two of these cases—from the FD administrative regions of Sobradinho and Lago Norte—were autochthonous, and 1 case resulted in death. VL cases have also been documented in domestic dogs in northern and southern areas of the FD, which has experienced unplanned urbanization of rural areas (Carranza-Tamayo et al. 2010).

An understanding of the distribution of phlebotomine species is essential for the prevention of leishmaniasis (Cutolo et al. 2013); however, knowledge on their geographic distribution in the FD is still poorly understood, and the phlebotomine fauna has rarely been sampled or compared to that of other regions. Studies analyzing the occurrence of sand flies in FD are scarce (Carvalho et al. 2010, Ferreira et al. 2015), which hinders the identification of areas at risk for *Leishmania* spp. transmission and epidemiological policies to prevent leishmaniasis. In order to update the list of phlebotomine species occurring in the FD, we studied the sand fly fauna from gallery forests.

Table 1. Institutional catalog number of the vouchers of phlebotomine species captured in this study.

Species (species code)	Catalog no. of the vouchers
Bichromomyia flaviscutellata (Bif)	CBRBif 1 – 632
	BNPBif 1 – 36
Brumptomyia brumpti (Brb)	ALFBrb 1 – 2
Brumptomyia guimaraesi (Brg)	ALFBrg 1 – 29
	BGBBrg 1 – 2
Brumptomyia sp. (Brsp)	ALFBrsp 1 – 41
Evandromyia bourrouli (Evb)	ALFEvb 1 – 7
	CBREvb 1 – 10
	BGBEvb 1
Micropygomyia longipennis (Mil)	ALFMil 1 – 3
	CBRMil 1 – 7
Micropygomyia ferreirana (Mif)	CBRMif 1 – 4
Micropygomyia sp. (Misp)	ALFMisp 1
Nyssomyia whitmani (Nyw)	CBRNyw 1 – 10
	BNPNyw 1 – 16
	BGBNyw 1
Psathytromyia abonnenci (Paab)	ALFPaab 1
	BNPBi 1 – 5
Psathyromyia aragaoi (Paar)	ALFPaar 1
Psathyromyia bigeniculata (Pab)	ALFPab 1 – 41
	BNPPab 1
	BGBPab 1
Psathyromyia lutziana (Pal)	ALFPal 1 – 17
	BNPPal 1
	BGBPal 1
Psathyromyia pradobarrientosi (Pap)	ALFPap 1 – 276
	CBRPap 1
	BNPPap 1 – 5
	BGBPap 1 – 3
Pintomyia christenseni (Pic)	ALFPic 1 – 26
Pintomyia kuscheli (Pik)	ALFPik 1
Pintomyia monticola (Pim)	ALFPim 1 – 24
Sciopemyia sordellii (Scs)	ALFPic 1 – 3

Methods

Study site. The Brazilian Federal District is located in the Center-West Region of Brazil. Its total area is 5,801,937 km^2, and it currently has a population of nearly 3 million inhabitants (IBGE 2016). The climate is classified as tropical according to the Köppen system, and it has 2 defined seasons: the rainy season (from October to April) and the dry season, which corresponds to the months of May to September (Klink and Machado 2005, Carvalho et al. 2010). The study was performed in four areas: the Brasília National Park (BNP), the Água Limpa Farm (ALF), the Contagem Biological Reserve (CBR), and the Botanical Garden of Brasília (BGB), which are shown in Figure 1.

BNP (15°41′51″S, 047°57′32W) is the most important protected area in the FD, occupying an area of 30,566 ha (Parna 1997). ALF (15°57′17″ S, 047°58′30″ W) is the University of Brasília farm where agricultural and ecological research projects are conducted. Its area is approximately 4340 ha, 3140 ha of which are protected areas and 1200 ha are used for production. The BGB (15°53′17″ S, 047°50′33″ W) occupies an area of 4518 ha, 526 ha of which are open to public visitation (Fonseca 2001). CBR (15°40′33 S, 047°51′45″ W) covers an area of 3460 ha. Two water catchment systems within the boundaries of this protected area contribute to the potable water supplies in FD. The region around the CBR has been undergoing an urban settlement process that has increased in the last 10 years.

Data collection. In each gallery forest, 2 trap lines of 1 km each were established (Fig. 1), with 20 HP light traps (Pugedo et al. 2005) set up. They were spaced 50 m apart and left for 4 consecutive days in May and September 2014. One Shannon trap was set 50 m away from each trapping line. The total capture effort included 1280 HP traps (160 per area in each month) and 16 Shannon traps (4h). The selection of these periods was based on previous studies, which reported a more frequent occurrence of Phlebotominae in the dry months in the FD (Ferreira et al. 2015).

All specimens captured were conditioned in a 70% ethanol solution. The specimens were dissected, and the head and genitalia were clarified and mounted in Canada balsam between the slide and the cover slip for species identification (Galati 2015). The remaining body parts (the thorax, part of the abdomen, wings, and legs) were placed in Eppendorf tubes containing phosphate buffered saline (PBS) (1 body part per tube). The tubes were labeled and placed in the freezer. Some of the specimens were identified only to the genus level due to the integrity of the material. Codes of each specimen collected were based on the names of the studies areas (ALF: Água Limpa Farm, CRB: Contagem Biological Reserve, BNP: Brasília National Park, BGB: Botanical Garden of Brasília) followed by abbreviations of the genera and of the species name.

Data analyses. The frequencies of traps that were positive and negative for sand flies in each gallery forest and month were compared using Fisher's exact test and a significance level of 5%. Relative species abundance was also calculated for the sand flies identified.

Results

The total phlebotomine sample from the 4 gallery forests in May and September 2014 consisted of 1209 specimens (615 males, 594 females) and 16 species. All vouchers were deposited in the Faculty of Medicine of the University of Brasilia (Table 1).

The greatest number of species was recorded in ALF and the greatest number of specimens was captured in CBR. Moreover, few individuals were captured in BNP and BGB (Table 2). Total capture success was 18%; the results were highest at the ALF ($p < 0.05$) in September ($p < 0.05$), as shown in Figure 2.

From the 472 phlebotomines captured at the ALF, 276 (58.5%) were identified as *Psathyromyia pradobarrientosi*, which was also the most frequent in May (90.3%). This species was followed by *Brumptomyia guimaraesi* and *Pa. bigeniculata*, both with 41 specimens

Table 2. Phlebotomine species captured in the 4 areas studied in the Brazilian Federal District, in May and September 2014. Total species richness in the gallery forests: 16 species.

Species	ALF			CBR			BNP			BGB			Total	%
	May	Sep	%	May	Sep	%	May	Sep	%	May	Sep	%		
Bichromomyia flaviscutellata	0	0	0	454	178	95.1	31	5	56.2	0	0	0	668	55.3
Brumptomyia brumpti	2	0	0.4	0	0	0	0	0	0	0	0	0	2	0.2
Brumpotomyia guimaraesi	0	29	6.1	0	0	0	0	0	0	0	2	22.2	31	3.6
Brumptomyia sp.	0	41	8.7	0	0	0	0	0	0	0	0	0	41	2.4
Evandromyia bourrouli	0	7	1.5	0	10	1.5	0	0	0	0	1	11.1	18	1.5
Mycropygomyia longipennis	1	2	0.6	0	7	1	0	0	0	0	0	0	10	0.8
Mycropygomyia ferreirana	0	0	0	0	4	0.6	0	0	0	0	0	0	4	0.3
Micropygomyia sp.	0	1	0.2	0	0	0	0	0	0	0	0	0	1	0.1
Nyssomyia whitmani	0	0	0	6	4	1.5	16	0	25	1	0	11.1	27	2.2
Psathyromyia abonnenci	1	0	0.2	0	0	0	5	0	7.8	0	0	0	6	0.5
Psathyromyia aragaoi	0	1	0.2	0	0	0	0	0	0	0	0	0	1	0.1
Psathyromyia bigeniculata	0	41	8.7	0	0	0	0	1	1.6	0	1	11.1	43	3.6
Pintomyia christenseni	0	26	5.5	0	0	0	0	0	0	0	0	0	26	2.2
Pintomyia kuscheli	0	1	0.2	0	0	0	0	0	0	0	0	0	1	0.1
Psathyromyia lutziana	10	7	3.6	0	0	0	1	0	1.6	1	0	11.1	19	1.3
Pintomyia monticola	1	22	4.9	0	0	0	0	0	0	0	0	0	23	1.9
Psathyromyia pradobarrientosi	100	176	58.5	1	0	0.2	5	0	7.8	1	2	33.3	285	23.6
Sciopemyia sordellii	1	2	0.6	0	0	0	0	0	0	0	0	0	3	0.2
Total	116	356		461	203		58	6		3	6		1.209	

ALF = Água Limpa Farm. CBR= Contagem Biological Reserve. BNP = Brasília National Park. BGB = Botanical Garden of Brasília.

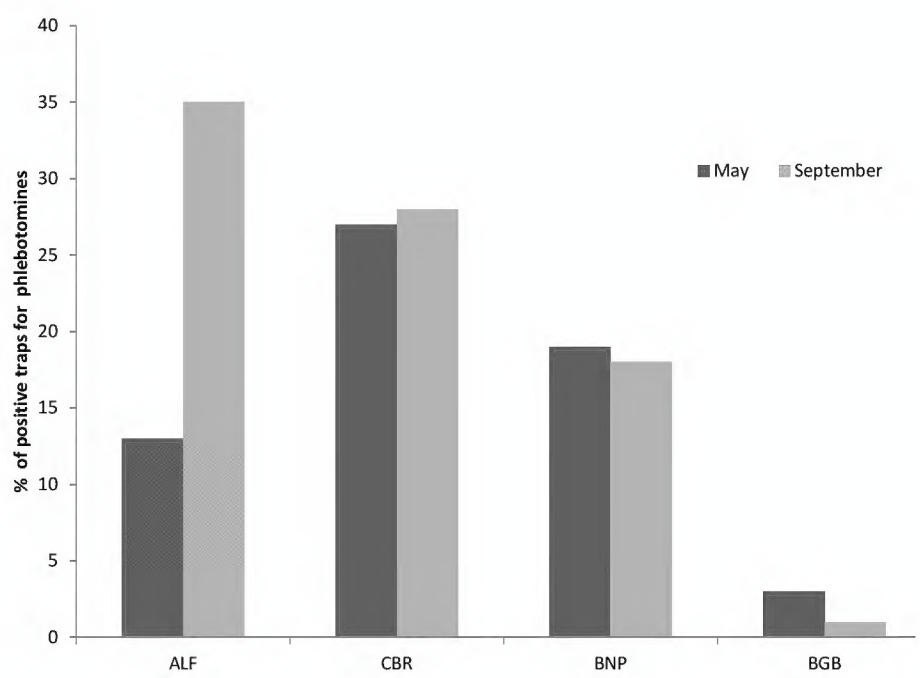

Figure 2. Percentage of positive traps for Phlebotominae in four gallery forests in Brazilian Federal District, in May and September 2014. ALF = Água Limpa Farm. CBR = Contagem Biological Reserve. BNP = Brasília National Park. BGB = Botanical Garden of Brasília.

each (11.5%). These species were particularly abundant in September (Table 2).

A total of 664 specimens were collected in the CBR. From these, *Bichromomyia flaviscutellata* was the most abundant both in May ($N = 454$; 98.4%) and September ($N = 178$; 87.7%). Low frequencies of *Nyssomyia whitmani* and *Evandromyia bourrouli* were observed (Table 2).

In BNP, a total of 64 phlebotomines were collected, 36 of which were *Bi. flaviscutellata* (56.25%). The second most common species in the BNP was *Ny. whitmani*, and overall, more species were collected in May than in September (Table 2). In the BGB, capture success was very low. Only 9 specimens were collected (Table 2).

This study found 5 new records: *Psathyromyia pradobarrientosi* is reported for the first time in Brazil and the Cerrado biome; *Brumptomyia guimaraesi*, *Br. brumpti*, *Mi. ferreirana*, and *Ev. bourrouli* are reported in the FD for the first time herein. In addition, 2 species known as vectors were collected in the gallery forests sampled: *Ny. whitmani* and *Bi. flaviscutellata*. The other species are not of any known epidemiological importance. This study has updated the list of sand fly fauna in the FD, where 35 species have now been recorded (Table 3).

Diagnoses of the newly recorded species.

Brumptomyia brumpti (Larrousse, 1920)

Material examined. Tables 1, 2; Figure 3a–c.

Male: superior proepimeral setae present; anepimeral setae absent; gonostyle with 2 external superior spines in the same tubercle; posterior spurs on the ascoids; ca 20 semi-foliaceous bristles on gonocoxite forming a tuft implanted directly on the structure (Fig. 3a); paramere not bifurcate and pilosous; aedeagus with conic format.

Brumptomyia guimaraesi (Coutinho & Barretto, 1941)

Material examined. Tables 1, 2; Figure 3a–c.

Male: superior proepimeral and anepimeral setae present (Fig. 3c); anepisternal inferior setae absent; gonostyle with 2 external superior spines on the same tubercle and presenting 2 apical spines; posterior spurs on ascoids; bristles on gonocoxite implanted in a compact and circular area (Fig. 3b); ejaculatory duct ca10× longer than ejaculatory pump.

Micropygomyia* (*Sauromyia*) *ferreirana (Barretto, Martins & Pellegrino, 1956)

Material examined. Tables 1, 2; Figure 4a–c.

Table 3. Updated list of phlebotomine species in the Brazilian Federal District. Total species richness: 35 species.

Family/ subfamily/tribe	SubTribe	Species	Habitat	Reference*
Psychodidae/ Phlebotominae/ Phlebotomini	Brumptomyiina	*Brumptomyia brumpti* (Larrousse 1920)[a]	Gallery forest	1
		Brumptomyia avellari (Costa Lima 1932)	—	3, 5
		Brumptomyia pintoi (Costa Lima 1932)	—	3, 6
		Brumptomyia guimaraesi (Coutinho & Barretto 1941)[a]	Gallery forest	1
	Sergentomyiina	*Micropygomyia longipennis* (Barretto 1946)	Gallery forest	1, 3, 5, 6
		Mycropygomyia ferreirana (Barretto, Martins & Pellegrino 1956)[a]	Gallery forest	1
		Mycropygomyia acanthopharynx (Martins, Falcão & Silva 1962)	—	3, 5, 6
		Mycropygomyia quinquefer (Dyar 1929)	Gallery forest	4
	Lutzomyiina	*Evandromyia bourrouli* (Barretto & Coutinho 1941)[a]	Gallery forest	1
		Evandromyia bacula (Martins, Falcão & Silva 1965)	—	3, 5, 6
		Evandromyia teratodes (Martins, Falcão & Silva 1964)	—	3, 5, 6
		Evandromyia sallesi (Galvão & Coutinho 1939)	—	3, 5, 6
		Evandromyia corumbaensis (Galati, Nunes, Oshiro & Rego 1989)	—	3, 6
		Evandromyia saulensis (Floch & Abonnenc 1944)	—	3, 5, 6
		Evandromyia evandroi (Costa Lima & Antunes, 1936)	—	3, 5, 6
		Evandromyia lenti (Mangabeira 1938)	—	3, 5, 6
		Pintomyia christenseni (Young & Duncan 1994)	Gallery forest	1, 3, 6
		Pintomyia kuscheli (Le Pont, Martinez, Torrez-Espejo & Dujardin 1998)	Gallery forest	1, 4
		Pintomyia monticola (Costa Lima 1932)	Gallery forest	1, 4
		Pintomyia fischeri (Pinto 1926)	—	3, 6
		Lutzomyia longipalpis (Lutz & Neiva 1912)	Peridomicile	3, 5, 6
		Lutzomyia ischnacantha Martins, Souza & Falcão 1962	—	3, 6
		Sciopemyia servulolimai (Damasceno & Causey 1945)	—	3, 6
		Sciopemyia sordellii (Shannon & Del Ponte 1927)	Gallery forest	1, 3, 4, 5
		Psathyromyia lutziana (Costa Lima 1932)	Gallery forest	1, 3, 4, 5, 6
		Psathyromyia pradobarrientosi (Le Pont, Matias, Martinez & Dujardin 2004)[a,b,c]	Gallery forest	1
		Psathyromyia brasiliensis (Costa Lima 1932)	—	3, 6
		Psathyromyia abonnenci (Floch & Chassignet 1947)[d]	Gallery forest	1, 4, 5
		Psathyromyia aragaoi (Costa Lima 1932)	Gallery forest	1, 3, 5, 6
		Psathyromyia bigeniculata (Floch & Abonnenc 1941)[d]	Gallery forest	1, 4, 5
	Psychodopygina	*Psychodopygus davisi* (Root 1934)	—	3
		Nyssomyia whitmani (Antunes & Coutinho 1939)	Gallery forest, peridomicile	1, 3, 4, 5, 6
		Nyssomyia neivai (Pinto 1926)	—	3, 5, 6
		Nyssomyia intermedia (Lutz & Neiva 1912)	—	2, 3, 6
		Bichromomyia flaviscutellata (Mangabeira 1942)	Gallery forest, peridomicile	1, 3, 4, 5, 6

*References: 1 = current study; 2 = Carvalho et al. (1989); 3 = Carvalho et al. (2010); 4 = Ferreira et al. (2014); 5 = Almeida et al. (2015); 6 = Galati (2015).
[a] First record in the Federal District.
[b] Species erroneously identified as *Pa. runoides* (see details in the text).
[c] First record in Brazil.
[d] Species erroneously identified as *Pa. shannoni*, voucher numbers: *Psathyromyia pradobarrientosi* # ALFPap 1 – 276, CBRPap 1, BNPPap 1 – 5, BGBPap 1 – 3; *Brumptomyia guimaraesi* # ALFBrg 1 – 29, # BGBBrg 1 – 2; *Brumptomyia brumpti* # ALFBrb 1 – 2; *Micropygomyia ferreirana* # CBRMif 1 – 4; *Evandromyia bourrouli* # ALFEvb 1 – 7, CBREvb 1 – 10, BGBEv 1. - no data about specific habitat. ALF = Água Limpa Farm. CBR= Contagem Biological Reserve. BNP = Brasília National Park. BGB = Botanical Garden of Brasília.

Female: Segment II of the palpus shorter than IV (Fig. 4a); setae on the katepisternum and post-alar setae absent; labial fork present; pharynx with developed spines (Fig. 4b); 2 pairs of posterior teeth in the cibarium (Fig. 4b); absence of sclerotinized points in cibarial camara; fagellomere III with same length as labrum–epipharynx; body of the spermathecae with 2 distinct rings (Fig. 4c).

Evandromyia* (*Evandromyia*) *bourrouli (Barretto & Coutinho, 1941)

Material examined. Tables 1, 2; Figure 5a–c.

Female: eyes longer than clypeus (Fig. 5a); mesonotum and pleura brown; common duct of spermathecae reaching middle of genital fork; body of spermathecae striate (Fig. 5b).

Male: gonocoxite with basal bristles (Fig. 5c); gonostyle with external spines isolated (Fig. 5c); lateral lobe with 3 spatulated setae (Fig. 5c); paramere not bifurcated.

Psathyromyia* (*Forattiniella*) *pradobarrientosi (Le Pont, Matias, Martinez & Dujardin, 2004)

Material examined. Tables 1, 2; Figure 6a–c.

Male: posterior spur on the ascoids (Fig. 6a); ejaculatory duct ca 3.2–5.7× longer than pump (Fig. 6b); gonostyle with external spine at its middle (Fig. 6b); ejaculatory

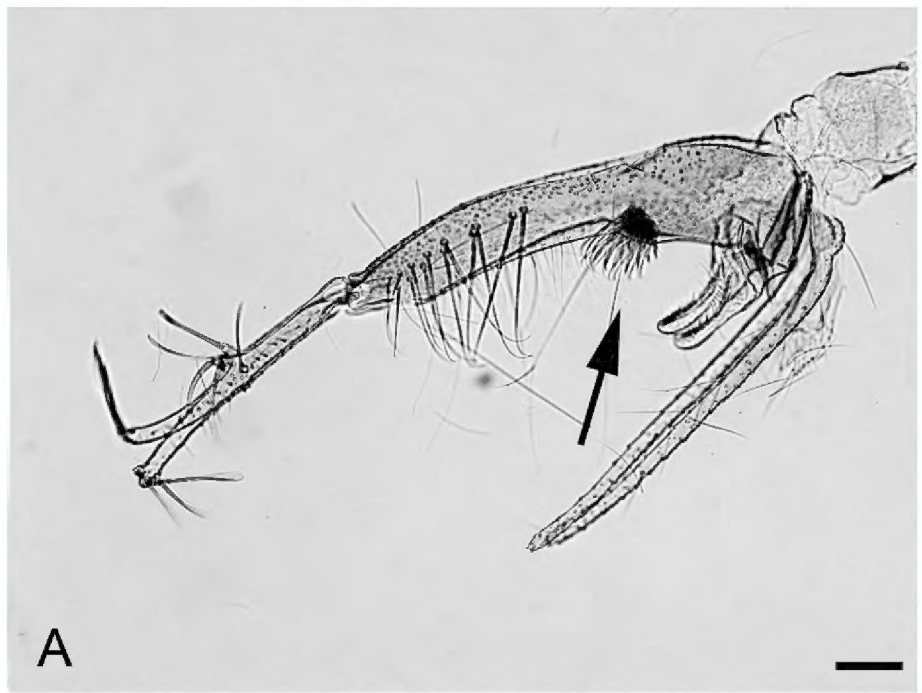

Figure 3. A. Male genitalia of *Brumptomyia brumpti*, voucher number ALFBrb 1; setae shows semi-foliaceous bristles on the gonocoxite forming a tuft implanted directly on the structure. **B.** Male genitalia of *B. guimaraesi*, voucher number ALFBrg 20; setae shows bristles on the gonocoxite implanted in a compact and circular area. **C.** Thorax of the male of *B. guimaraesi*; voucher number ALFBrg 20; setae shows superior proepimeral and anepimeral setae present; in the detail is the anepimeral region highlighted. Scale bars = 100 µm.

duct with losangular tip (Fig. 6c) and 2× the width of the anterior portion; aedeagus short.

Discussion

This study contributes to the knowledge of the phlebotomine fauna in the gallery forests of FD, a region which has been considered endemic for VL since 2005. Between 2010 and 2015, the number of species found in the FD increased to 35, 5 of which were recorded for the first time in the current study.

Because of urban expansion, deforestation has been the main factor affecting environmental changes in the FD. In the CBR, residences are located close to the remaining fragments of the Cerrado and even of the gallery forests. As a result, there is interaction between wild fauna and humans, as well as between these sylvatic species and domestic animals. These confined animals become a new, easy, and safe blood source for sand flies. The finding of species of epidemiological importance in the gallery forest of the BNP (and the presence of *Ny. whitmani* in particular) means that extra attention is required in the completion of the urban development project involving the creation of a new neighborhood in the northwestern sector of the FD. The ecological impact of human activity on the area may be reflected in the increased number of CL cases in dogs and humans in the area (Cardoso et al. 2015).

The main neighborhoods that share borders with the BGB, which are known as Lago Sul and Jardim Botânico, also experience strong human pressure due to overcrowding and urbanization in the Cerrado that surrounds the region. The occurrence of phlebotomine sand flies in BGP represents a risk of the possible expansion of leishmaniasis to humans and domesticated animals in the region.

The ALF exhibited the greatest species richness. Among the species found in the ALF, *Pa. pradobarrientosi* was the most frequently captured. This species was found in the other 3 study areas as well. *Pa. pradobarrientosi* was also the most abundant species in the BGB. The male of *Pa. pradobarrientosi* had been described previously from the Bolivian Amazon during the rainy season. The occurrence of this species in Brazilian FD (in all areas sampled in the current study) may be associated with the presence of forest connections between Brazilian Cerrado and Bolivian Amazon. The gallery forests may function as a refuge for many phlebotomine species and as ecological corridors in which these insects move along the riverbanks, as has been suggested in studies on other faunal groups (Moraes et al. 2015). It is a species morphologically similar to *Pa. runoides*, so it was long confused with it (Le Pont et al. 2004). Ferreira et al. (2015) recorded *Pa. runoides* in FD, but after reviewing the slides, one of us (AJA), based in Galati's taxonomic proposal observed a misidentification. The description of the female is still being developed (AJA, personal communication).

Sábio et al. (2014, 2016a, b) reviewed the taxonomic status of the species within the subgenus *Psathyromyia* Barretto, 1962 showing that *Pa. shannoni* does not occur in Brazil. The species *Pa. shannoni*, *Pa. abonnenci*, *Pa. limai*, *Pa. bigeniculata*, *Pa. pifanoi*, *Pa. baratai* and *Pa. ribeirensis* are still included in a taxonomic category named shannoni series (Sábio et al. 2016b) due the aspect of the spermathecae of the females. Ferreira et al. (2015) recorded *Pa. shannoni* in the FD, but based in the

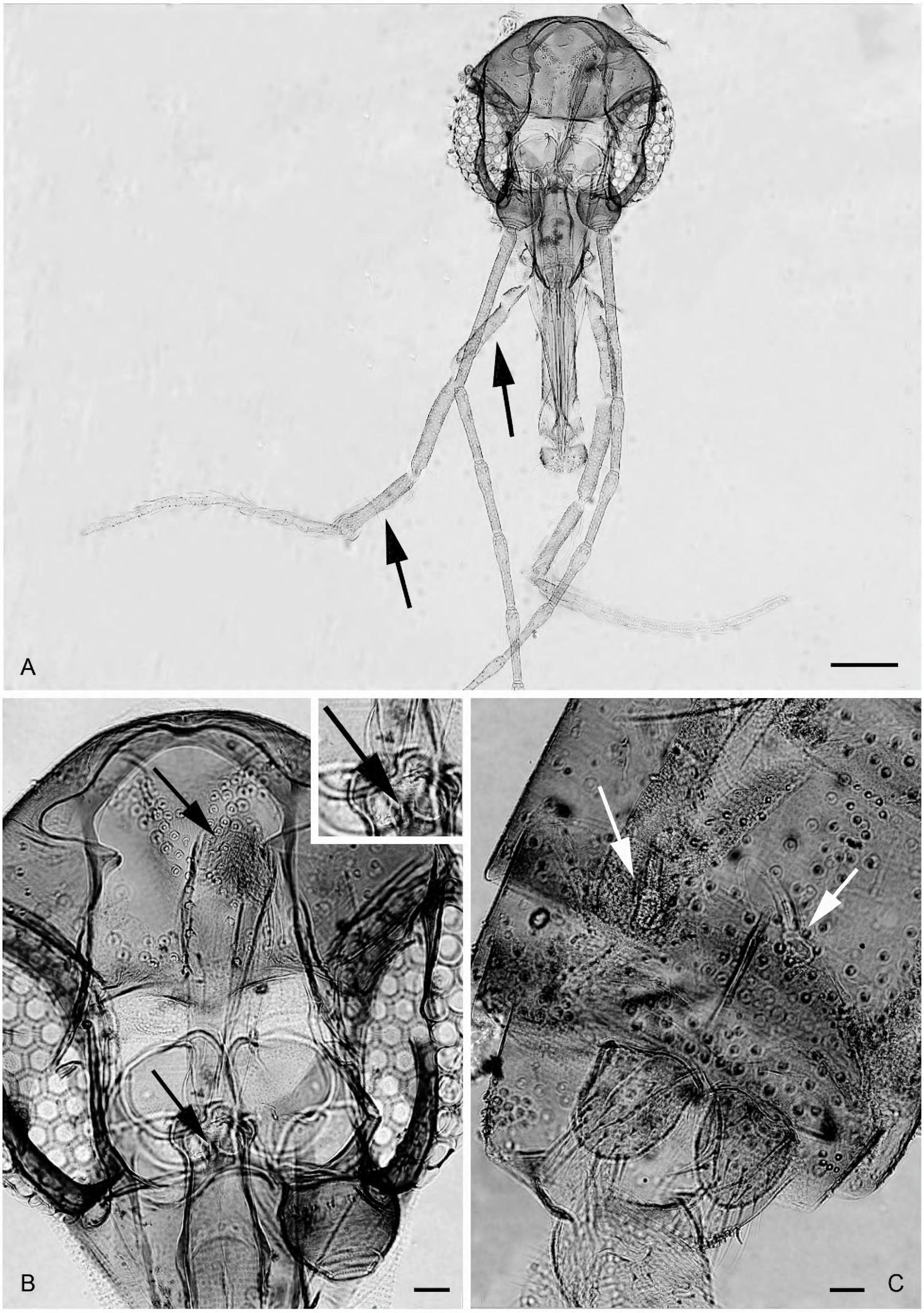

Figure 4. A. Female head of *Micropygomyia ferreirana*, voucher number CBRMif 1; setae shows the relationship between II segment of palpus (basal setae) and IV segment of palpus (proximal setae). **B.** Pharynx and cibarium of the female of *M. ferreirana* voucher number RM247; setae shows the spines in the pharynx and the teeth of cibarium; in the detail cibarium is highlighted. **C.** Spermathecae of *M. ferreirana*, voucher number CBRMif 1; setae shows the head of the structure. Scale bars: A = 100 µm; B, C = 20 µm.

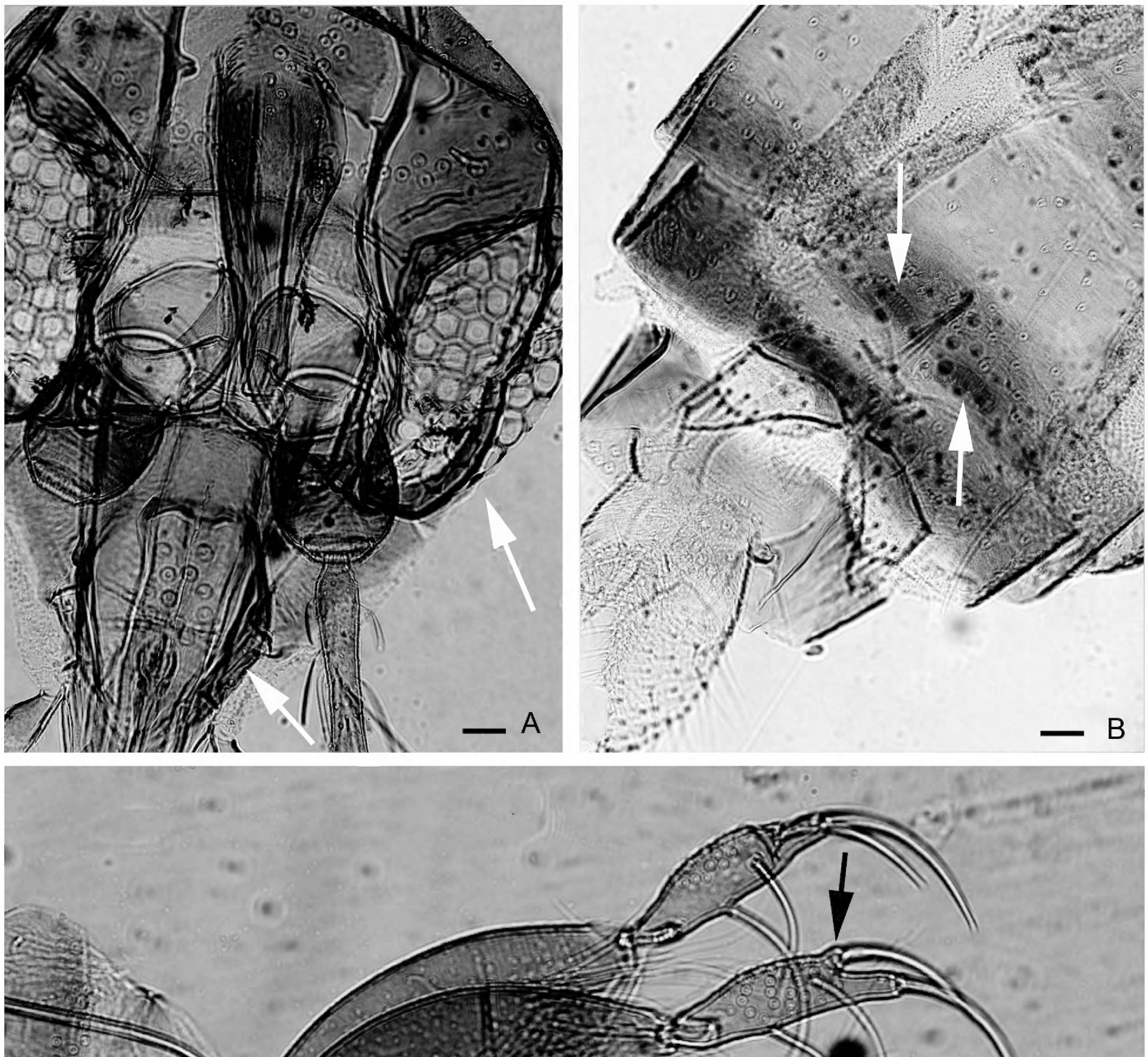

Figure 5. **A.** Pharynx and cibarium of the female of *Evandromyia bourrouli*, voucher number ALFEvb 1; setae shows the relationship between the eye and the clypeus. **B.** Spermathecae of *E. bourrouli*, voucher number ALFEvb 2; setae shows the body of the structure. **C.** Male genitalia of *E. bourrouli*, voucher number ALFEvb 7; setae shows the bristles of the gonocoxite and the lateral lobe with three spatulated setae. Scale bars: A, B = 20 µm; C = 50 µm.

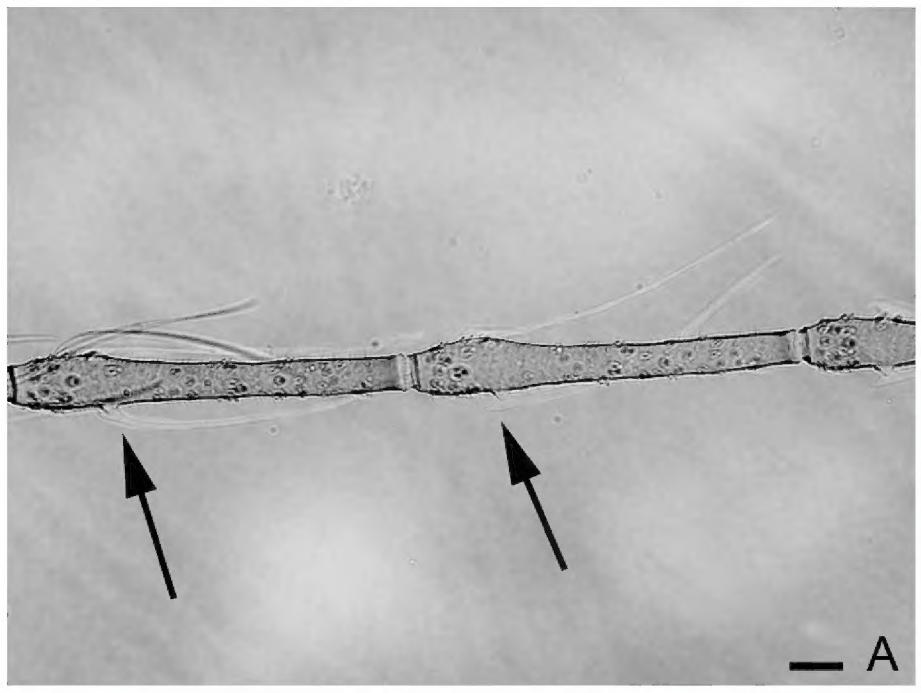

Figure 6. A. Ascoid in flagellomeres of *Psathyromyia pradobarrientosi*, voucher number BNPPap 1; setae shows the ascoids. **B.** Male genitalia of *P. pradobarrientosi*, voucher number ALFPap 1; setae shows the relationship between the pump and the ejaculatory duct and the external spine in the middle of the gonostyle. **C.** Tip of the ejaculatory duct of *P. pradobarrientosi*, voucher number ALFPap 1; setae shows the losangular format of the structure. Scale bars: A, B = 20 µm; C = 100 µm.

key published by Sábio et al. (2014) and Galati (2015) it was possible to revise the identification of the species previously identified as *Pa. shannoni*. *Psathyromyia bigeniculata* was a junior synonym of *Pa. shannoni* that was revalidated by Sábio et al. (2014) and correctly identified in the present study. Specimens of *Pa. bigeniculata* can be distinguished of *Pa. abonnenci* based in the thoracic coloration and males can be also separated using the distribution of the bristles on the dorsal margin of the paramere.

Brumptomyia guimaraesi specimens were collected in the ALF and the BGB, while *Brumptomyia brumpti* was only collected in the ALF. *Brumptomyia* species are not important in the eco-epidemiology of leishmaniasis (Eckert and Souza 2010), and according to Aguiar and Medeiros (2003), these species suck blood from armadillos (Edentata, Dasypodidae). The insects are always found on fallen leaves on the forest floor and in mammal dens (Eckert and Souza 2010, Cutolo et al. 2013).

Four *Mycropygomyia ferreirana* specimens were found in the CBR in September and in the Center-West Region of Brazil was recorded only in Mato Grosso do Sul (Galati 2015). Some species of the genus *Micropygomyia*, subgenus *Sauromyia*, such as *Mi. quinquefer* (Paiva et al. 2010), *Mi. ferreirana* (Rocha et al. 2010, Rêgo et al. 2015), *Mi. capixaba*, *Mi. villelai*, and *Mi. peresi* (Rêgo et al. 2015) have been recorded infected by *Leishmania* species through molecular analyzes in some states of Brazil. According to Aguiar and Medeiros (2003), species included in these genera bite cold-blooded animals, but as mentioned by Rêgo et al. (2015) the finding of *Leishmania* DNA in this genus suggests that these sand flies have fed on hosts susceptible to parasite infection.

Evandromyia bourrouli, a species inhabiting forests and armadillo dens (Dorval et al. 2010), was captured only in September. Dorval et al. (2010) found a high density of this species in the state of Mato Grosso do Sul using Disney traps. According to their study, almost all females were found engorged, which seemed to reflect their affinity for rodents as a source of blood. They also noted that this species has been collected in forest areas and human environments, particularly those in the Center-West and Northeastern regions of Brazil. Araújo-Pereira et al. (2014) found *Ev. bourrouli* in forests in the Brazilian state of Acre, and Vilela et al. (2013) reported this species in rural and periurban areas in Guaraí (Tocantins state). *Evandromyia bourrouli* has been found in many different habitats ranging from preserved forests to animal shelters (chicken coops and pigpens), which reflects their eclectic behavior. However, this species has not been reported as a vector of pathogens to humans.

The predominance of *Bi. flaviscutellata* was expected in forested areas as reported by Carvalho et al. (2010), who compared sand flies captures in households, peridomestic environments, and gallery forests. The current study found many specimens of *Bi. flaviscutellata*, particularly in the CBR and the BNP. Given the presence of small mammals infected by *Leishmania* (*Leishmania*) *amazonensis* in these areas (Cardoso et al. 2015), this species may be responsible for the enzootic circulation of this parasite, which is an etiological agent of diffuse cutaneous leishmaniasis.

Similarly, *Ny. whitmani* was captured in the CBR and the BNP, as well as in the BGB. Carvalho et al. (2010) found the species in all the locations and habitats studied in the FD (households, peridomestic environments, and

gallery forests). This finding is indicative of the potential degree of domiciliation of this species. It reflects the importance of this species as a *Leishmania braziliensis* vector in the FD. *Ny. whitmani* is also considered the most important CL vector in many parts of Brazil (Costa et al. 2007).

Psathyromyia pradobarrientosi, Pa. lutziana, Mi. longipennis, Sciopemyia sordellii, and *Pintomyia monticola* were found in the ALF in May and September. In both months, *Pa. pradobarrientosi* was found in the BGB, *Bi. flaviscutellata* and *Ny. whitmani* were found in the CBR, and *Bi. flaviscutellata* was found in the BNP. The other species were found to be distributed irregularly between these 2 months. These monthly differences in abundance and occurrence indicate that these species respond differently depending on the presence of different blood-based food sources, breeding grounds, shelters, and the season. However, the small area of influence of light traps is important; the attraction radius for phlebotomines (small insects) is not more than 5 m (Dye et al. 1991). The success of samplings, therefore, depends on the displacement of species within the geographical space studied (where the traps are distributed).

No *Lu. longipalpis* specimens were captured in the gallery forests studied. This species is currently listed as the main VL vector in the Americas. Initially, VL was associated with rural habitats and the outskirts of large cities; however, this profile has changed in certain regions where the disease has been found to be urbanized. This process has been associated with environmental modifications caused by human activity, by the rapid migratory process, by the interactions between wild reservoirs and infected dogs and their mobilization into areas without transmission, and by the adaptation of the vector *Lu. longipalpis* to peridomestic environments (Vilela et al. 2013).

Changes in behavior exhibited by *Lu. longipalpis* enable this species to adapt to increasing losses in biodiversity. Moreover, the frequent exposure to insecticides in urban areas rendered them resistant. These changes may explain population increases of this species and the consequent propagation of the disease to peridomestic and domestic habitats (Aguiar et al. 2014, Salomón et al. 2015). The absence of this species in the gallery forests sampled reflects its adaptive nature, which may be reflected in its apparently opportunistic behavior in relation to sources of carbohydrates and blood (Brazil 2013). This is especially likely, given its presence in peridomestic and domestic habitats in the FD (Carvalho et al. 2010). From an evolutionary perspective, this behavior may have favored its vector competence in urban areas (Brazil 2013).

Little is known about the sand fly fauna in the FD relative to other regions in Brazil. Carvalho et al. (2010) listed 27 phlebotomine species in different habitats in the FD. The current study has found 35 species in FD. This number of species is representative of the region, given the fact that 47 species were reported in Goiás, which is approximately 59 times the size of the FD (Almeida et al. 2015). Future studies with efforts focused on sampling methods similar to those of the current research and on other vegetation types may be able to reveal a larger species richness in the FD. Relevant information on species richness, abundance, and occurrence at different periods of the year may be used to better understand the epidemiology of diseases and to increase knowledge available on phlebotomine behavior in the gallery forests close to the urban areas of the FD.

Acknowledgements

The authors would like to thank William Quatman, who reviewed the English text and the students of Veterinary Medicine Débora Juliana Rodrigues Martins, Jéssica Toledo de Almeida, Cristiana Santiago Nery de Souza, Agda Regina Melo Silva, Thamerson Felipe Lelis dos Santos, Saemis Ferreira de Souza, Tâmela Stéfany Lopes Silva, Renê Cunha de Castro, Rubens Geraldo de Souza, Jadson José Gomes da Silva, the students of Biology Sciences Lana Cristina Evangelista Ferreira Sá, Hayana Maria Boaventura Costa, Tauana de Souza Ferreira and Mr. Barbosa José Bezerra for their fundamental collaboration with the fieldwork. The fieldwork was approved by Instituto Chico Medes de Biodiversidade (ICMBio), authorization no. 36808-6 and by all the conservation units involved.

Authors' Contributions

AR collected the data, processed the material, and wrote the text and tables. AJA collected the data, identified the specimens, wrote the diagnoses, and coordinated the work. DAR collected the data and photographed the material. JCBF collected the data. RVT processed the material. MTO collected the data and RGG collected and analyzed the data, prepared the figures, and coordinated the work. All authors reviewed the manuscript.

References

Aguiar GM, Medeiros WM (2003) Distribuição regional e hábitats das espécies de flebotomíneos do Brasil. In: Rangel EF, Lainson R (Eds) Flebotomíneos do Brasil. Editora FIOCRUZ, Rio de Janeiro, 207–255.

Aguiar GM, Azevedo ACR, Medeiros WM, Alves JRC, Rendeiro V (2014) Aspects of the ecology of phlebotomines (Diptera: Psychodidae: Phlebotominae) in an area of cutaneous leishmaniasis occurrence, municipality of Angra dos Reis, Cost of Rio de Janeiro State, Brazil. Revista do Instituto de Medicina Tropical de São Paulo 56 (2): 143–149. https://doi.org/10.1590/S0036-46652014000200010

Almeida PS, Andrade AJ, Sciamarelli A, Raizer J, Menegatti JA, Hermes S, Carvalho MSL, Gurgel-Gonçalves R (2015) Geographic distribution of phlebotomine sandfly species (Diptera: Psychodidae) in Central-West Brazil. Memórias do Instituto Oswaldo Cruz 110 (4): 551–559. https://doi.org/10.1590/0074-02760140462

Alvar J, Vélez ID, Bern C, Herrero M, Desjeux P, Cano J (2012) Leishmaniasis worldwide and global estimates of its incidence. PLoS ONE 7 (5): e35671. http://.doi.org/10.1371/journal.pone.0035671

Antunes PCA, Coutinho JO (1939) Notas sôbre Flebotomos Sul-Americanos. II. Descrição de *Flebotomus whitmani* sp. e da armadura

bucal de algumas espécies. Boletim Biológico (n.s.) 4 (3): 448–453.

Araujo-Pereira T, Fuzari AA, Andrade Filho JD, Pita-Pereira DD, Britto C, Brazil RP (2014) Sand fly fauna (Diptera: Psychodidae: Phlebotominae) in an area of leishmaniasis transmission in the municipality of Rio Branco, state of Acre, Brazil. Parasites and Vectors 7: 360. https://.doi.org/10.1186/1756-3305-7-360

Barretto MP, Coutinho JO (1941) Contribuição ao conhecimento dos flebótomos de São Paulo. VI. Descrição de duas novas espécies (Diptera, Psychodidae). Papéis Avulsos do Departamento de Zoologia 1 (24): 223–236.

Barretto MP (1946) Uma nova espécie de flebótomo do Estado de Goiás, Brasil, e chave para determinação das espécies afins (Diptera, Psychodidae). Revista Brasileira de Biologia 6 (3): 427–434.

Barretto MP, Martins AV, Pellegrino J (1956) Notas sôbre flebotomíneos do nordeste do Estado de Minas Gerais, com as descrições de duas espécies novas (Diptera, Psychodidae). Folia Clinica et Biologica 26: 47–56.

Brazil RP (2013) The dispersion of *Lutzomyia longipalpis* in urban areas. Revista da Sociedade Brasileira de Medicina Tropical 46 (3): 263–264. https://doi.org/10.1590/0037-8682-0101-2013

Cardoso RM, Araújo NNSL, G.S. Romero GS, Minuzzi-Souza TTC, Dietrich AG, Mendes JD, Reis ML, Ferreira JBC, Hecht MM, Gurgel-Gonçalves R (2015) Expanding the knowledge about *Leishmania* species in wild mammals and dogs in the Brazilian savannah. Parasites & Vectors 8: 171. https://doi.org/10.1186/s13071-015-0780-y

Carranza-Tamayo CO, Carvalho MSL, Bredt A, Bofill MIR, Rodrigues RMB, da Silva AD, Cortez SMF, Romero GAS (2010) Autochthonous visceral leishmaniasis in Brasília, Federal District, Brazil. Revista da Sociedade Brasileira de Medicina Tropical 43 (4): 396–399. https://doi.org/10.1590/S0037-86822010000400012

Carvalho LSM, Bredt A, Meneguin SRE, Oliveira C (2010) Flebotomíneos (Diptera: Psychodidae) em áreas de ocorrência de leishmaniose Tegumentar americana no Distrito Federal, Brasil, 2006 a 2008. Epidemiologia e Serviços de Saúde 19 (3): 227–237. https://doi.org/10.5123/S1679-49742010000300005

Carvalho LSM, Lusíosa ES, Naves HAM (1989) Contribuição ao conhecimento da fauna flebotomínica do Estado de Goiás e Distrito Federal. II 1986–1987. Revista de Patologia Tropical 18 (1): 7–14. https://doi.org/10.5216/rpt.v18i1.20212

Costa Lima A, Antunes PCA (1936) Sôbre um novo phlebótomo encontrado no Norte do Brasil. Brazil Médico 50: 419–422.

Costa Lima A (1932) Sôbre os phlebótomos americanos (Diptera, Psychodidae). Memórias do Instituto Oswaldo Cruz 26 (1): 15–69.

Costa SM, Cechinel M, Bandeira V, Zannuncio JC, Lainson R, Rangel EF (2007) *Lutzomyia* (*Nyssomyia*) *whitmani* l. (Antunes & Coutinho, 1939) (Diptera: Psychodidae: Phlebotominae): geographical distribution and the epidemiology of American cutaneous leishmaniasis in Brazil mini-review. Memórias do Instituto Oswaldo Cruz 102 (2): 149–153. https://doi.org/10.1590/S0074-02762007005000016

Coutinho JO, Barretto MP (1941) Contribuição ao conhecimento dos flebótomos de São Paulo. VII. Descrição de *Phlebotomus guimaraesi* sp. e das fêmeas de *Phlebotomus pestanai* Barretto e Coutinho, 1941e de *Phlebotomus lanei* Barretto e Coutinho, 1941 (Diptera, Psychodidae). Revista de Biologia e Higiene 11 (11): 74–88.

Cutolo AA, Galati BAE, VonZuben JC (2013) Sandflies (Diptera, Psychodidae) from forest areas in Botucatu municipality, central western São Paulo State, Brazil. Journal of Venomous Animals and Toxins including Tropical Diseases 19: 15. https://doi.org/10.1186/1678-9199-19-15

Damasceno RG, Causey OR (1945) Estudo sôbre Flebotomus no Vale Amazônico. Parte III. Descrição de *F.servulolimai* e *F.wilsoni* (Diptera, Psychodidae). Memórias do Instituto Oswaldo Cruz 42 (3): 635–644.

Dias SE, Falcão LA, Silva EJ, Martins VA (1989) Redescrição do macho e descrição da fêmea de *Lutzomyia* (Helcocyrtomyia) *ferreirana* (Barreto, Martins & Pellegrino, 1958) (Diptera, Psychodidae, Phlebotominae). Memórias do Instituto Oswaldo Cruz 84 (2): 245–248.

Dorval MEC, Alves TP, Cristaldo G, Rocha HC, Alves MA, Oshiro ET, Oliveira AG, Brazil RP, Galati EAB, Cunha RV (2010) Sand fly captures with Disney traps in area of occurrence of *Leishmania* (*Leishmania*) *amazonensis* in the State of Mato Grosso do Sul, mid-western Brazil. Revista da Sociedade Brasileira de Medicina Tropical 43: 491–495. https://doi.org/10.1590/S0037-86822010000500003

Dyar HG (1929) The present knowledge of the American species of *Phlebotomus rondani* (Diptera, Psychodidae). American Journal of Tropical Medicine and Hygiene 10 (1): 112–124.

Dye C, Davies CR, Lainson R (1991) Communication among phlebotomine sand flies: a field study of domesticated *Lutzomyia longipalpis* populations in Amazonian Brazil. Animal Behavior 42: 183–192. https://doi.org/10.1016/S0003-3472(05)80549-4

Eckert J, Souza GD (2010) Flebotomíneos (Diptera: Psychodidae: Phlebotominae) no município de Estrela e primeiro registro de *Lutzomyia pascalei* (Coutinho & Barretto) no Rio Grande do Sul. Revista Brasileira de Biociências 8 (4): 399–402.

Ferreira JBC, Macedo MA, Rocha DA, Ferreira TS, Obara MT, Gurgel-Gonçalves R (2015) Ocorrência de flebotomíneos (Diptera: Psychodidae) em matas de galeria no Distrito Federal, Brasil. EntomoBrasilis 7 (3): 216–221. https://doi.org/10.12741/ebrasilis.v7i3.398

Floch H, Abonnenc E (1944) Phlébotomes de la Guyane française. VII. Description de six espèces nouvelles. Institut Pasteur de la Guyane et du Territoire de l'Inini 80: 1–20.

Floch H, Abonnenc E (1941) Phlébotomes de la Guyane Française (II). Institute Pasteur Guyane Territoire l'Inini 28: 1–16.

Floch H, Chassignet R (1947) Phlébotomes de la Guyane Française (XXI). Institute Pasteur Guyane Territoire l'Inini 157: 1–3.

Fonseca FO (Ed.) (2001) Olhares sobre o Lago Paranoá. Secretaria de Meio Ambiente e Recursos Hídricos, Brasília, 425 pp.

Galati EAB, Nunes VLB, Oshiro ET, Rego FA Jr (1989) Nova espécie de Phlebotominae, *Lutzomyia corumbaensis*, sp. (Diptera, Psychodidae) do complexo *Lutzomyia cortelezzii*. Revista Brasileira de Entomologia 33 (3): 465–475.

Galati EAB (2015) Phlebotominae (Diptera, Psychodidae) Classificação, morfologia, terminologia e identificação de Adultos. Apostila Disciplina HEP 5752 Bioecologia e Identificação de Phlebotominae Vol. I. Universidade de São Paulo, São Paulo. http://www.fsp.usp.br/egalati. Accessed on: 2015-11-30.

Galati EAB, Nunes VLB, Dorval MEC, Oshiro ET, Cristaldo GE, Marcos A, Rocha HC, Garcia WB (1996) Estudo dos flebotomíneos (Diptera, Pychodidae) em área de leishmaniose tegumentar no Estado de Mato Grosso do Sul, Brasil. Revista de Saúde Pública 30 (2): 115–128. https://doi.org/10.1590/S0034-89101996000200002

Galvão ALA, Coutinho JO (1939) Contribuição ao estudo dos Flebotomus do Estado de São Paulo. *Flebotomus sallesi*, n. sp. (Diptera, Psychodidae): Anais da Faculdade de Medicina da Universidade de São Paulo 15: 125–139.

IBGE (2016) Tabelas de estimativas de população. http://www.ibge.gov.br/home/estatistica/populacao/estimativa2016/estimativa_tcu.shtm. Accessed on: 2017-8-8.

Klink CA, Machado RB (2005) Conservation of Brazilian Cerrado. Conservation Biology 19: 707–713. http://doi.org/10.1111/j.1523-1739.2005.00702

Larrousse F (1920) Nouvelle espèce américaine du genre *Phlebotomus*, *Phlebotomus brumpti*, sp. nov: Bulletin de la Société de Pathologie Exotique 12: 659–662.

Le Pont F, Martinez EM, Torres M, Dujardin JP (1998) Description de 5 nouveles spéces de *Lutzomyia* (Diptera: Psychodidae) de région Subandine. Bulletin de la Société Entomologique de France 103: 159–173.

Le Pont F, Matias A, Mariinez E, Dujardin JP (2004) Phlébolomes de Bolivie: description de quatre nouvelles espèces de *Lutzomyia* (Diptera, Psychodidae). Bulletin de la Société Entomologique de France 109 (2): 145–154.

Lutz A, Neiva A (1912) Contribuição para o conhecimento das espécies do gênero *Phlebotomus* existentes no Brasil. Memórias do Instituto Oswaldo Cruz 4 (1): 84–95.

Mangabeira Filho O (1942) Descrição dos machos de 24 novas espécies: 7ª contribuição ao estudo dos flebotomus. Memórias do Instituto Oswaldo Cruz 37: 259–387.

Mangabeira O (1938) Sobre duas novas espécies de *Phlebotomus* (Diptera: Psychodidae). Memórias do Instituto Oswaldo Cruz 33: 349–356.

Martins AV, Falcão AL, Silva JE (1962) Nota sôbre os flebótomos do Estado de Goiás, com a descrição de duas espécies novas e da fêmea de *Lutzomyia longipennis* (Barretto, 1946) e a redescrição do macho da *L.evandroi* (Costa Lima & Antunes, 1936) (Diptera, Psychodidae). Revista Brasileira de Malariologia e Doenças Tropicais 14 (4): 379–399.

Martins AV, Falcão AL, Silva JE (1964) Um novo flebótomo do Estado de Goiás, *Lutzomyia teratodes* sp.n. (Diptera, Psychodidae). Revista Brasileira de Biologia 24 (3): 321–324.

Martins AV, Falcão AL, Silva JE (1965) Notas sobre flebótomos do Território de Rondônia, com a descrição de seis espécies novas (Diptera, Psychodidae). Revista Brasileira de Biologia 25 (1): 1–20.

Martins AV, Souza MA, Falcão AL (1962) Estudos sobre os flebótomos do Estado de Minas Gerais. IV: Descrição de duas novas espécies do sub-gênero "*Lutzomyia*" França, 1924 (Diptera, Psychodidae). Revista Brasileira de Biologia 22 (1): 39–46.

Moraes JLP, Santos CLC, Bastos VJC, Pereira ACN, Moraes LS, Moraes JR, Costa MC, Rebêlo JMM (2015) Espécies de flebotomíneos (Diptera, Psychodidae) em matas ciliares na transição entre a Amazônia úmida e o Nordeste semi-árido do Brasil. Entomotrópica 30 (3): 20–29.

Paiva BR, Oliveira AG, Dorval MEMC, Galati EAB, Malafronte RS (2010) Species-specific identification of *Leishmania* in naturally infected sand flies captured in Mato Grosso do Sul state, Brazil. Acta Tropica 115: 126–130.

Parna (1997) Plano de Manejo do Parque Nacional de Brasília. Convênio IBAMA x FUNATURA. Encartes: 4–6. http://www.icmbio.gov.br/portal/images/stories/imgs-unidades-coservacao/PARNA%20Brasilia.pdf. Accessed on: 2016-5-23.

Pinto C (1926) *Phlebotomus neivai* e *Phlebotomus fischeri* n.sp. Sôbre o aparelho espicular dos phlebotomos e seu valor especifico. Sciencia Medica 4 (7): 370–375.

Pugedo H, Barata RA, França-Silva JC, Silva JC, Dias ES (2005) HP: um modelo aprimorado de armadilha luminosa de sucção para a captura de pequenos insetos. Revista da Sociedade Brasileira de Medicina Tropical 38: 70–72. https://doi.org/10.1590/S0037-86822005000100017

Rêgo FD, Rugani JMN, Shimabukuro PHF, Tonelli GB, Quaresma PF, Gontijo CMF (2015) Molecular detection of *Leishmania* in phlebotomine sand flies (Diptera: Psychodidae) from a cutaneous leishmaniasis focus at Xakriabá Indigenous Reserve, Brazil. PLoS ONE 10 (4): e0122038. https://doi.org/10.1371/journal.pone.0122038

Rocha LS, Falqueto A, Santos CB, Ferreira AL, Graça GC, Grimaldi-Jr G, Cupollilo E (2010) Survey of natural infection by *Leishmania* in sandfly species collected in southeastern Brazil. Transactions of the Royal Society of Tropical Medicine and Hygiene 104: 461–466.

Root FM (1934) Some American species of *Phlebotomus* with short terminal palpal segments. American Journal of Hygiene 20: 233–246.

Sábio PB, Andrade AJ, Galati EAB (2014) Assessment of the taxonomic status of some species included in the *Shannoni* complex, with the description of a new species of *Psathyromyia* (Diptera: Psychodidae: Phlebotominae). Journal of Medical Entomology 51 (2): 331–341. https://doi.org/10.1603/ME13153

Sábio PB, Andrade AJ, Galati EAB (2016a) Description of *Psathyromyia (Psathyromyia) baratai* sp. n. (Diptera: Psychodidae: Phlebotominae) from Cantareira State Park, São Paulo, Brazil. Journal of Medical Entomology 53: 83–90. https://doi.org/10.1093/jme/tjv173

Sábio PB, Brilhante AF, Quintana MG, Andrade AJ, Galati EAB (2016b) On the synonyms of *Psathyromyia (Psathyromyia) shannoni* (Dyar, 1929) and *Pa. bigeniculata* (Floch & Abonnenc, 1941) and the resuscitation of *Pa. pifanoi* (Ortiz, 1972) with the description of its female (Diptera: Psychodidae: Phlebotominae). Journal of Medical Entomology 53 (5): 1140–1147. https://doi.org/10.1093/jme/tjw094

Salomón OD, Feliciangeli MD, Quintana MG, Afonso MMS, Rangel EF (2015) *Lutzomyia longipalpis* urbanisation and control. Memórias do Instituto Oswaldo Cruz 110 (7): 831–846. https://doi.org/10.1590/0074-02760150207

Sampaio RNR, Gonçalves MC, Leite VA, França BV, Santos G, Carvalho MSL, Tauil PL (2009) Study on the transmission of American cutaneous leishmaniasis in the Federal District. Revista da Sociedade Brasileira de Medicina Tropical 42: 686–690. https://doi.org/10.1590/S0037-86822009000600015

Sampaio RNR, Rocha RAA, Marsden PD, Cuba CC, Barreto AC (1980) Leishmaniose tegumentar americana. Casuística do Hospital Escola da UnB. Anais Brasileiros de Dermatologia 55: 69–76.

Shannon RC, Del Ponte E (1927) Cuatro notas sobre especies nuevas de dipteros nematóceros, hematófagos de la República Argentina. Revista del Instituto Bacteriológico 4 (1): 724–726.

Shimabukuro PHF, Andrade AJ, Galati EAB (2017) Checklist of American sand flies (Diptera, Psychodidae, Phlebotominae): genera, species, and their distribution. ZooKeys 660: 67–106. https://doi.org/10.3897/zookeys.660.10508

Shimabukuro PHF, Galati EAB (2011) Checklist dos Phlebotominae (Diptera, Psychodidae) do Estado de São Paulo, Brasil, com comentários sobre sua distribuição geográfica. Biota Neotropica 11 (1a): 1–20.

Vilela ML, Pita-Pereira D, Azevedo CG, Godoy RE, Britto C, Rangel EF (2013) The phlebotomine fauna (Diptera: Psychodidae) of Guaraí, state of Tocantins, with an emphasis on the putative vectors of American cutaneous leishmaniasis in rural settlement and periurban áreas. Memórias do Instituto Oswaldo Cruz 108 (5): 578–585. https://doi.org/10.1590/S0074-02762013000500007

Young DG, Duncan MA (1994) Guide to the identification and geographic distribution of *Lutzomyia* sand flies in Mexico, the West Indies, Central and South America (Diptera: Psychodidae). Memoirs of the American Entomological Institute 54: 1–881.